

Optimizing cardiac energy substrate metabolism: a novel therapeutic intervention for ischemic heart disease

Jagdip S. Jaswal, Virgilio J.J. Cadete and Gary D. Lopaschuk

Cardiovascular Research Group, Departments of Pediatrics and Pharmacology, Faculty of Medicine and Dentistry, University of Alberta, Edmonton, Alberta, T6G 2S2, Canada

Correspondence: Dr Gary D. Lopaschuk, 4–23 Heritage Medical Research Centre, University of Alberta, Edmonton, Alberta, Canada T6G 2S2.
E-mail: gary.lopaschuk@ualberta.ca

Abstract

Ischemic heart diseases, encompassing and ranging from angina pectoris to acute myocardial infarction, have a major impact on both cardiac energy metabolism and cardiac function. In the normal heart, energy metabolism and function are exquisitely matched. However, during and after ischemia there are both a decrease in energy production and disturbances in the balance between use of fatty acid and of glucose by the heart. The dominance of fatty acid oxidation as a source for the generation of ATP at the expense of glucose oxidation during and after ischemia has a negative impact on both cardiac efficiency and cardiac contractile function. Thus optimizing energy substrate metabolism, such that the efficiency of both generating and utilizing ATP is maximized has emerged as a novel therapeutic intervention in various manifestations of ischemic heart disease. For example, the antianginal benefit of trimetazidine can be attributed to the partial inhibition of fatty acid oxidation and the reciprocal increase in glucose oxidation. This optimization of the balance between fatty acid and glucose metabolism results in an improvement in the efficiency of both the generation and utilization of ATP. Other pharmacological agents also exploit this plasticity and interdependence between the pathways of fatty acid and glucose oxidation. This is achieved either by altering flux through these metabolic pathways, or by altering the availability of circulating energy substrates. Thus the multitude of targets available to optimize myocardial energy metabolism may significantly increase the armamentarium of therapeutic interventions for preserving cardiac contractile function and limit the untoward effects of ischemic heart disease.

■ *Heart Metab.* 2008;38:5–14.

Introduction

Alterations in myocardial energy substrate metabolism contribute significantly to ischemic heart disease. Angina pectoris is a common form of ischemic heart disease, and has an impact on both the amount of energy produced by the heart and the type of fuel it metabolizes. In this context, a growing body of evidence indicates that the modulation and optimizing of

myocardial energy substrate metabolism are useful therapeutic interventions for the treatment of various forms of ischemic heart disease, including angina pectoris.

The heart is an omnivorous organ. It uses fatty acids, glucose, lactate, and ketone bodies as fuels to sustain contractile function. The contribution of each substrate to the overall production of ATP is tightly regulated, with each pathway possessing a considerable

Basic article

Jagdip S. Jaswal, Virgilio J. J. Cadete and Gary D. Lopaschuk

degree of plasticity and interdependence. Under normal aerobic conditions, the heart relies primarily on fatty acids as substrates for oxidative metabolism. Fatty acid β -oxidation normally contributes 60–70% of total ATP production in the healthy adult heart; the remainder is provided mainly by carbohydrate oxidation (glucose oxidation and lactate oxidation), and also (at a very low percentage) by the oxidation of ketone bodies [1,2]. With respect to the major ATP-producing processes in the heart, fatty acid oxidation produces more ATP per molecule oxidized than does glucose oxidation; however, fatty acid oxidation requires a greater amount of oxygen per molecule of ATP produced. Thus fatty acid oxidation is less efficient than glucose oxidation with regards to ATP production per molecule of oxygen consumed.

Disease states and other conditions (eg, elective cardiac surgical procedures) that result in a serious insult to the heart can perturb the tightly regulated energetic balance in the heart, which can contribute to myocardial damage. An example of this is ischemic heart disease, which dramatically alters both the rate of energy production and the source of energy supply. During ischemia, oxygen availability is reduced as a result of deficient tissue perfusion, resulting in a mismatch between oxygen demand and oxygen supply. A decrease in oxygen supply results in a concomitant decline in the rates of mitochondrial oxidative metabolism. During ischemia, glycolysis becomes increasingly important because of its ability to generate ATP in the absence of oxygen. Unfortunately, this can lead to the intracellular accumulation of lactate and protons (H^+), which in itself can decrease cardiac efficiency. Furthermore, during ischemia, plasma free fatty acid concentrations increase dramatically, and result in the rapid recovery of fatty acid oxidation during subsequent reperfusion of the ischemic myocardium. These increased rates of fatty acid oxidation uncouple glycolysis and glucose oxidation, and so increase H^+ production. The dramatic increase in rates of fatty acid oxidation in early reperfusion can impair the recovery of cardiac function.

As knowledge of how cardiac energy metabolism is regulated increases, the potential application of metabolic modulation to the treatment of ischemic heart disease has become the subject of extensive research and review. The aim of this article is to present the mechanistic basis for the use of pharmacological agents to optimize myocardial energy substrate metabolism in order to limit the deleterious consequences of ischemia.

Myocardial energy metabolism

Under aerobic conditions, more than 50% of the ATP produced in the heart is derived from mitochondrial

oxidative phosphorylation (Figure 1) [1,2]. Reducing equivalents (H^+ and electrons) are transferred from substrates to the mitochondria by the reduced forms of flavine adenine dinucleotide ($FADH_2$) and nicotinamide adenine dinucleotide (NADH), generated by dehydrogenase reactions occurring during β -oxidation, the Krebs (tricarboxylic acid [TCA]) cycle and pyruvate oxidation (glucose oxidation). The extents to which the various metabolic pathways contribute to the production of ATP are dependent on energetic demand, which itself is determined by contractile work.

In the presence of a normal oxygen supply, glucose and fatty acids both undergo oxidation through different processes that link at the level of the TCA cycle

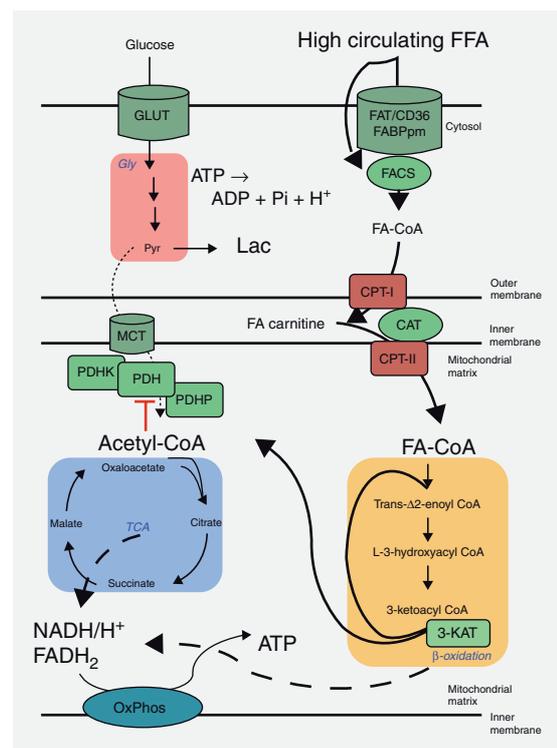


Figure 1. Glucose/fatty acid cycle. In the presence of high concentrations of circulating free fatty acids, fatty acid uptake and oxidation increase considerably, resulting in an accumulation of acetyl coenzyme A (CoA), which in turn inhibits the pyruvate dehydrogenase (PDH) complex, uncoupling glycolysis from subsequent oxidation. This further results in the hydrolysis of glycolytic ATP uncoupled from oxidative metabolism and concomitantly increased production of cytosolic protons, which can result in intracellular acidosis. CAT, carnitine acyl translocase; CPT, carnitine palmitoyl transferase; FABPpm, fatty acid binding protein of the plasma membrane; FACS, fatty acyl CoA synthase; FA CoA, fatty acyl CoA; FAT/CD36, fatty acid transporter; FFA, free fatty acid; GLUT, glucose transporter; Gly, glycolysis; 3-KAT, 3-ketoacyl CoA thiolase; Lac, lactate; MCT, monocarboxylic transporter; OxPhos, oxidative phosphorylation; PDHK, pyruvate dehydrogenase kinase; PDHP, pyruvate dehydrogenase phosphatase; Pi, inorganic phosphate; Pyr, pyruvate; TCA, tricarboxylic acid cycle.

Basic article

Metabolic modulation for ischemic heart disease

(Figure 1). The presence of this common pathway is central both to the mechanisms regulating flux through these pathways and to the interdependence of these processes for ATP production.

Glucose used for the generation of ATP originates from the blood stream, or is liberated from endogenous glucose stores (ie, glycogen). Glucose enters the cardiac myocyte via glucose transporters (GLUTs) [3]. GLUT 4 is the main myocardial glucose transporter, and is sensitive to insulin stimulation, whereas a small percentage of glucose transport occurs via the insulin-insensitive transporter, GLUT 1. By the glycolytic pathway, glucose is converted into pyruvate with the net production of two molecules of ATP and two molecules of NADH. In the presence of oxygen, pyruvate is oxidized (glucose oxidation) by the pyruvate dehydrogenase (PDH) complex to form acetyl coenzyme A (CoA), which then feeds into the TCA cycle. Alternatively, in the absence of adequate oxygen, pyruvate can be converted to lactate by the enzyme lactate dehydrogenase (LDH), to regenerate the NAD^+ required to maintain glycolysis. The PDH complex is rate-limiting for glucose oxidation, and is highly sensitive to product inhibition by acetyl CoA. When high rates of fatty acid oxidation are present, there is an increase in the concentration of acetyl CoA, which in turn can inhibit glucose oxidation [4–6]. This reciprocal inter-regulatory relationship between glucose oxidation and fatty acid oxidation was originally described by Philip Randle, and is known as the glucose/fatty acid cycle or Randle cycle [7].

On the other side of oxidative metabolism lies fatty acid oxidation. Fatty acid oxidation occurs mainly in the mitochondrial matrix and is highly dependent on the delivery of fatty acids, first from the plasma to the cytoplasm, and subsequently from the cytoplasm to the mitochondrial matrix. Fatty acids enter the cardiac myocyte either by passive diffusion or via protein-mediated uptake. The key transporters involved in fatty acid uptake are fatty acyl translocase (FAT/CD36) and the plasma membrane isoform of fatty acid binding protein (FABPpm) [3,8]. Fatty acids are then esterified to fatty acyl CoA, which is mediated by a family of fatty acyl CoA synthase (FACS) enzymes. The mitochondrial uptake of fatty acyl CoAs is mediated by carnitine palmitoyl transferases (CPT) I and II and carnitine acyl translocase (CAT) [1,9]. CPT-I is present on the outer mitochondrial membrane. It binds to fatty acyl CoAs and catalyzes the formation of fatty acyl carnitines which are transported to the mitochondrial inter-membrane space. There, CAT translocates fatty acyl carnitines into the matrix (in exchange for carnitine), where CPT-II re-esterifies acyl carnitines into acyl CoAs (Figure 1). Matrix acyl CoAs can then be progressively metabolized by fatty acid oxidation. Four main enzyme classes are involved in the mitochondrial fatty acid oxidation: acyl CoA de-

hydrogenase, 2-enoyl CoA hydratase, 3-hydroxyacyl CoA dehydrogenase and 3-ketoacyl CoA thiolase (3-KAT). In the fatty acid oxidation spiral, fatty acyl CoAs are broken down to acetyl CoA, which feeds into the TCA cycle for the production of ATP. Both acyl CoA dehydrogenase and 3-hydroxyacyl CoA dehydrogenase are sensitive to the redox state of the matrix (FAD/FADH_2 and NAD^+/NADH ratios). In the presence of high rates of glucose oxidation, the concentration of NADH is increased, and the redox state of the mitochondria favors an inhibition of fatty acid oxidation. Fatty acid oxidation is also regulated at the level of 3-KAT, which is sensitive to the acetyl CoA/CoA ratio, and in the presence of high glucose oxidation rates acetyl CoA accumulates and inhibits 3-KAT.

On the basis of the enzymes involved in fatty acid oxidation, there are numerous targets available that can be exploited to optimize and modulate myocardial energy metabolism in order to limit the untoward effects of ischemic heart disease. Several pharmacological approaches to the optimization of energy substrate metabolism at the level of the balance between fatty acid and glucose metabolism (Figure 2) are considered below.

Carnitine palmitoyl transferase I inhibitors

Carnitine palmitoyl transferase-I is considered to be the rate-limiting enzyme for mitochondrial uptake of fatty acids. As a result, pharmacological agents exerting their anti-ischemic effects by inhibiting CPT-I have potential for therapeutic use in the treatment of ischemic heart disease (Figure 2). CPT-I inhibitors that have been developed for this purpose include oxfenicine, etomoxir, and perhexiline. Several experimental studies have demonstrated that the protective effects of oxfenicine [10,11], etomoxir [12,13], and perhexiline [11,14] are associated with a shift in energy substrate metabolism from fatty acid oxidation towards glucose oxidation. Of these compounds, perhexiline has received the most clinical attention.

Perhexiline was frequently prescribed as an anti-anginal agent in the 1970s; however, its use declined in the 1980s because of adverse effects, including hepatic toxicity (steatosis and necrosis), and peripheral neuropathy [15]. The mechanism responsible for these adverse effects of perhexiline is believed to be accumulation of phospholipid, secondary to the inhibition of CPT-I, and these adverse effects can therefore be presumed to be shared by CPT-I inhibitors as a class of drugs. It is of importance to note that the hepatotoxic effects of perhexiline arise as a result of the inhibition of hepatic CPT-I [16]; however, in-vitro studies indicate that the cardiac isoform of CPT-I is more sensitive to inhibition by perhexiline than is the hepatic CPT-I isoform [17]. Furthermore, monitoring

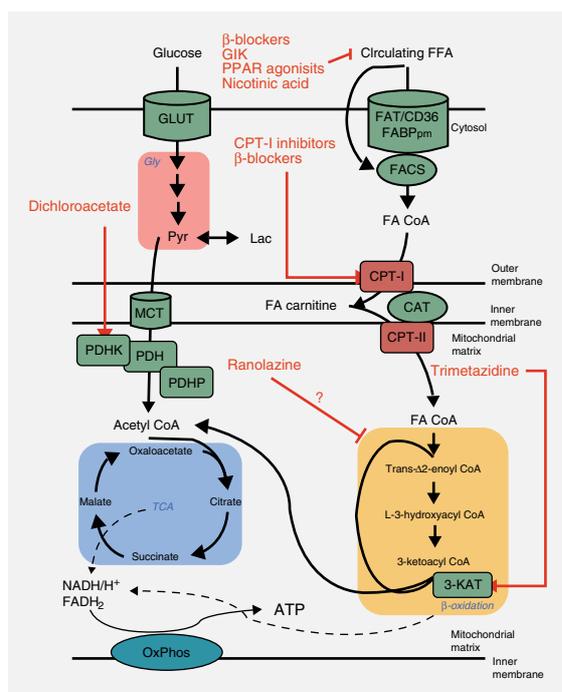


Figure 2. Pharmacological optimization of fatty acid and glucose metabolism. Three major metabolic pathways provide the necessary energy for heart function: fatty acid β -oxidation, glycolysis (Gly), and glucose oxidation (glycolysis + pyruvate [Pyr] oxidation by the tricarboxylic acid [TCA] cycle). Pharmacological compounds (shown in red) can modify energy substrate metabolism by modulating circulating concentrations of free fatty acids (FFA) and enzymatic activity at the levels of glucose oxidation, carnitine palmitoyl transferase (CPT)-I, and fatty acid oxidation. CAT, carnitine acyl translocase; CoA, coenzyme A; FABPpm, fatty acid binding protein of the plasma membrane; FA CoA, fatty acyl CoA; FACS, fatty acyl CoA synthase; FAT/CD36, fatty acid transporter; FFA, free fatty acid; GLUT, glucose transporter; Gly, glycolysis; 3-KAT, 3-ketoacyl CoA thiolase; Lac, lactate; MCT, monocarboxylic transporter; OxPhos, oxidative phosphorylation; PDH, pyruvate dehydrogenase (complex); PDHK, pyruvate dehydrogenase kinase; PDHP, pyruvate dehydrogenase phosphatase.

the plasma concentration of perhexiline and maintaining it in the therapeutic range of 150–600 $\mu\text{g/L}$ markedly limits the serious adverse effects of the drug while preserving its anti-ischemic efficacy [18]. Accordingly, the potential of targeting myocardial energy substrate metabolism to limit the consequences of ischemia has led to a resurgence in the use of perhexiline.

With the biochemical mechanisms responsible for both its therapeutic and its adverse effects being elucidated, perhexiline is used as antianginal agent in New Zealand, Australia, and most European countries, on a named-patient basis. Furthermore, clinical trials have demonstrated the utility of perhexiline in refractory angina pectoris [18], aortic stenosis [19], and chronic heart failure (of ischemic and non

ischemic origin) [20], in which it improves symptomatic status, left ventricular ejection fraction, and quality of life. Therefore, inhibition of CPT-I and fatty acid oxidation, with the resultant reciprocal increase in glucose oxidation, is a cardioprotective strategy that is effectively utilized in diverse forms of ischemic heart disease.

β -Adrenoceptor antagonists and myocardial energy substrate metabolism

The anti-ischemic properties of β -adrenoceptor antagonists (β -blockers) are classically attributed to an oxygen-sparing effect elicited by negative inotropic and chronotropic actions. In addition to effects on cardiac contractility, β -blockers also possess additional anti-ischemic mechanisms related to energy substrate metabolism. They reduce neurohormonal activation, and thereby can reduce catecholamine-induced lipolysis, and hence circulating plasma fatty acid concentrations (Figure 2) – a major determinant of the rates of fatty acid oxidation. Furthermore, several clinical studies have indicated that β -blockers decrease fatty acid uptake [21,22] and increase left ventricular function, independently of decreased oxygen consumption [23,24] – effects indicative of increased cardiac efficiency (work/oxygen consumed). These effects are probably related to the ability of β -blockers to inhibit the activity of CPT-I (Figure 2) [25], and induce a shift in energy substrate metabolism from fatty acid oxidation towards glucose oxidation [26].

3-Ketoacyl-coenzyme A thiolase inhibitors

Being a key enzyme in fatty acid oxidation, 3-KAT has emerged as a target for modifying fatty acid oxidation. Trimetazidine is the first of a class of partial fatty acid oxidation inhibitors that competitively inhibit the terminal enzyme of fatty acid oxidation, long-chain 3-ketoacyl CoA thiolase (Figure 2) [27,28]. It is clinically utilized as an antianginal agent throughout Europe and more than 90 countries worldwide [29]. The protective effects of trimetazidine are demonstrable in experimental models of ischemia-reperfusion. It decreases ischemic contracture, and lessens the increase in diastolic pressure during reperfusion after ischemia [30], and inhibits cardiac myocyte apoptosis to preserve cardiac function during reperfusion [31]. With regards to the mechanism of its anti-ischemic action, trimetazidine also protects hearts from the deleterious effects of fatty acids on the recovery of cardiac function [32]. By virtue of inhibiting fatty acid oxidation, it reciprocally stimulates glucose oxidation [27,28], and thus improves the

Basic article

Metabolic modulation for ischemic heart disease

coupling between glycolysis and glucose oxidation, thereby decreasing the rate of H^+ production attributable to the hydrolysis of glycolytically derived ATP. These effects of trimetazidine on the pathways of fatty acid and glucose metabolism can prevent deleterious alterations in intracellular ionic homeostasis by diminishing the potential for intracellular acidosis during ischemia, in addition to intracellular Na^+ and Ca^{2+} overload during reperfusion [33,34]. Therefore, the metabolic effects of trimetazidine increase cardiac efficiency by sparing ATP hydrolysis from being utilized for correcting ionic homeostasis, thus increasing the amount of ATP hydrolysis available to drive meaningful contractile work.

Numerous clinical trials have demonstrated the efficacy of trimetazidine in various forms of ischemic heart disease, ranging from angina pectoris to acute myocardial infarction, ischemic cardiomyopathy, and heart failure. With respect to angina, the beneficial effects of trimetazidine include an increased time to 1 mm ST-segment depression, a reduction in weekly consumption of nitrates, and a reduction in the number of weekly angina attacks both in patients who have not undergone revascularization and in those undergoing revascularization via coronary artery bypass grafting procedures or percutaneous coronary intervention [35]. Trimetazidine also has cardioprotective effects in the setting of acute myocardial infarction, where it reduces reperfusion arrhythmias [36] and reduces the time to resolution of ST-segment elevation [37]. Furthermore, trimetazidine added to existing treatment has been shown to improve New York Heart Association (NYHA) functional class, decrease left ventricular end-systolic volume, and increase left ventricular ejection fraction in patients with ischemic cardiomyopathy and heart failure [38,39]. Therefore, the partial inhibition of fatty acid β -oxidation with trimetazidine limits the deleterious consequences of myocardial ischemia in varied manifestations of ischemic heart disease.

Ranolazine, similar to trimetazidine, is a partial inhibitor of fatty acid oxidation, although the molecular target responsible for this effect remains to be identified (*Figure 2*). It is now approved for use as an antianginal agent in the USA [40], and, in addition to its metabolic effects, appears to have antiarrhythmic activity. Experimental studies have demonstrated that ranolazine preserves mitochondrial ultrastructure, decreases tissue Ca^{2+} content, and improves the recovery of ventricular function during reperfusion after ischemia [41], in addition to decreasing the incidence of ventricular fibrillation during reperfusion [42]. It has also been reported to reduce the magnitude of myocardial stunning [43] and to reduce infarct size [44]. Furthermore, in canine models of heart failure, ranolazine improves ejection fraction and stroke volume without increasing oxygen consump-

tion, and hence increases myocardial efficiency [45,46].

Several of the beneficial actions of ranolazine are most probably attributable to either a greater amount of ATP synthesized at a given level of oxygen consumption or a more effective use of the energy released from ATP hydrolysis – effects that can be explained by a shifting of myocardial energy substrate preference from fatty acid oxidation to glucose oxidation (described above) [47–49].

Clinical trials have demonstrated the antianginal efficacy of ranolazine. Both as monotherapy and in combination therapy with standard antianginal regimens, it has been shown to increase exercise duration, increase time to 1 mm ST-segment depression, reduce the weekly number of angina attacks, and reduce weekly consumption of nitroglycerin [50–53]. Furthermore, the antianginal efficacy of ranolazine is similar in both non diabetic and diabetic patients [54].

Recent clinical trials have also indicated that ranolazine decreases the incidence of ventricular tachycardia, supraventricular tachycardia, and ventricular pauses, and has no adverse effect on survival, thus confirming its long-term safety and efficacy [55,56]. The antiarrhythmic effects of the drug may be attributable to its effects on various cardiac ionic currents, particularly the late Na^+ current (for review see [57]). Interestingly, the antianginal and antiarrhythmic effects of ranolazine occur at similar therapeutic concentrations (10–20 $\mu\text{mol/L}$), and thus are probably not mutually exclusive. The effects on ranolazine on the pathways of fatty acid and glucose oxidation probably underlie its antianginal activity by improving the efficiency of ATP production.

Dichloroacetate

Dichloroacetate, like trimetazidine and ranolazine, also facilitates the shift in the balance of myocardial energy substrate metabolism away from fatty acid oxidation towards glucose oxidation; however, unlike trimetazidine and ranolazine, dichloroacetate exerts direct effects on the mitochondrial PDH complex. It inhibits the activity of PDH kinase, and thus stimulates glucose oxidation (*Figure 2*). Experimental studies have demonstrated the efficacy of dichloroacetate in enhancing the recovery of cardiac function during reperfusion after ischemia both *in vitro* and *in vivo* [58–60]. The protective effects of dichloroacetate are accompanied by a stimulation of the rate of glucose oxidation, and a resultant improved coupling between glycolysis and glucose oxidation (which reduces the production of H^+ attributable to glucose metabolism), and an increase in myocardial efficiency (work per molecule of oxygen consumed) [61,62].

Although clinical experience with dichloroacetate is limited, its metabolic mechanism of action appears to persist in the setting of heart failure. In a small clinical trial, dichloroacetate increased left ventricular stroke volume and myocardial efficiency, effects accompanied by increased utilization of lactate [63], which itself is probably the result of an increased rate of pyruvate oxidation (ie, glucose oxidation). These metabolic effects of dichloroacetate may be of therapeutic relevance in angina pectoris; however, they remain to be assessed in this setting.

Glucose–insulin–potassium for acute coronary syndromes

The concept of using glucose–insulin–potassium (GIK) solutions to protect the ischemic myocardium in acute coronary syndromes encompassing clinical conditions ranging from myocardial infarction to unstable angina was initially introduced by Sodi-Pollares et al [64]. The beneficial effects of GIK on cardiac energy metabolism that underlie the protection it affords were originally proposed by L.H. Opie as promotion of glycolysis and reduction in circulating fatty acids (*Figure 2*), with a resultant decrease in cardiac fatty acid metabolism [65]. Indeed, experimental studies have demonstrated the ability of GIK to suppress circulating fatty acid concentrations, while maintaining circulating glucose concentration [66]. These effects on circulating energy substrate concentrations are effective in inducing a shift in myocardial substrate preference from fatty acid to glucose utilization [67], and in improving post-ischemic recovery of contractile function, reducing the release of creatine kinase and lactate dehydrogenase, and reducing infarct size [66,67]. However, the protective effects of GIK are not unambiguous, as previous reports also indicate a lack of reduction in infarct size [68]. This ambiguity of GIK in experimental studies may be related to the complex nature of its effects on myocardial energy substrate metabolism – specifically, its ability to accelerate the rate of glycolysis disproportionately to that of glucose oxidation, and thus to increase the rate of H^+ production attributable to myocardial glucose metabolism, which can impair postischemic recovery of function [69].

The ambiguous effects of GIK on myocardial protection are transferred to the clinical setting, where GIK has been shown to be beneficial, neutral, and detrimental. A meta-analysis of GIK in the ‘prethrombolytic’ era demonstrated the ability of GIK to reduce the mortality associated with acute myocardial infarction [70], which was also evident in the thrombolytic era, as demonstrated by the Diabetic Patients with

Acute Myocardial Infarction (DIGAMI) study [71] and the Estudios Cardiológicos Latinoamerica (ECLA) Collaborative Group [72]. However, a Polish GIK study (Pol-GIK) failed to demonstrate any benefit of GIK on cardiovascular mortality [73]. Furthermore, in contrast to the Dutch Glucose–Insulin–Potassium Study 1 (GIPS 1), which demonstrated a survival benefit of GIK in a subset of patients [74], the Dutch GIPS 2 study, which assessed the effects of GIK on mortality and infarct size had to be stopped early because of a potentially greater mortality in the GIK group [75]. These differences in clinical outcomes may be related to the differing doses used, the timing of administration, and the functional NYHA classes of the patient populations. As there remains no clear consensus with regards to the beneficial, neutral, or detrimental clinical effects of GIK in acute coronary syndromes, further study is warranted.

Peroxisome proliferator-activated receptor agonists

Peroxisome proliferator-activated receptors (PPARs) are members of the ligand-activated nuclear hormone receptor superfamily. PPARs exert major influences on lipid metabolism at the whole-body level (*Figure 2*); specifically, they have a central role in regulating the balance between fatty acid oxidation and fatty acid storage, by regulating the expression of enzymes involved in both fatty acid oxidation and lipogenesis [76]. Three distinct PPAR isoforms, with overlapping yet differing/preferential tissue distributions, have been identified in mammals: PPAR α , PPAR γ , and PPAR δ . Expression of PPAR α is greatest in tissues with a high capacity for fatty acid oxidation, including the liver, skeletal muscle, and heart [76,77]. PPAR γ is expressed to the greatest extent in adipose tissue, with only low concentrations detected in the skeletal and cardiac muscle. PPAR δ is the predominant isoform expressed in skeletal muscle, white adipose tissue, and brown adipose tissue (in rodents) [78]. Importantly, PPARs are the molecular targets of several clinically useful pharmacological agents that alter whole-body lipid metabolism.

PPAR α is the molecular target of the antihyperlipidemic fibrate class of drugs (clofibrate, fenofibrate, gemfibrozil). Fibrates differentially affect the fatty acid binding capacity of cytosolic proteins from different tissues. They increase the fatty acid binding capacity of liver and kidney while not affecting that of skeletal muscle and decreasing that of cardiac cytosolic proteins [79]. This may be associated with the ability of fibrates to increase the expression and activity of long-chain acyl CoA synthetase in extracardiac tissue [80]. Furthermore, fibrates have been shown to increase the expression of the enzymes involved in fatty acid

Basic article

Metabolic modulation for ischemic heart disease

oxidation preferentially in the liver [81]. Taken together, these effects increase extracardiac fatty acid utilization, decreasing both circulating plasma and myocardial fatty acid concentrations, and thus myocardial fatty acid oxidation. Experimental studies have demonstrated that fibrates can improve the recovery of cardiac function after ischemia [82], and reduce infarct size [83] – protective effects that may arise as a result of the partitioning of fatty acids away from the heart.

PPAR γ is the molecular target of the oral antidiabetic thiazolidinedione class of drugs (rosiglitazone, pioglitazone, troglitazone). Thiazolidinediones prevent the spillover of lipids from adipose tissue into non adipocytes (eg, cardiac myocytes), thereby increasing adiposity, an effect attributed to a reduction in the ectopic deposition of fatty acids in tissues not suited for the storage of excess lipid. Experimental studies have demonstrated that thiazolidinediones can decrease circulating plasma triglyceride [84] and fatty acid [84,85] concentrations, while increasing both myocardial glucose uptake [86] and net lactate uptake [85] (indicative of increased glucose oxidation) and glucose oxidation [84]. These alterations in energy substrate selection and metabolism translate into improvements in the recovery of cardiac function after ischemia [84,85,87].

Interestingly, there exists discordance between the protective effects of thiazolidinediones in experimental studies and the potential to exacerbate the symptoms of heart failure in some diabetic patients treated with these compounds. Specifically, thiazolidinediones can cause fluid retention and peripheral edema, effects that are of concern in patients with heart failure [88]. Furthermore, the findings of a recent meta-analysis indicated that the use of thiazolidinediones in patients with type 2 diabetes mellitus is associated with an increase in the risk of myocardial infarction and an increased risk of death from cardiovascular causes [89]. Therefore, despite the potentially beneficial effects of thiazolidinediones on circulating plasma fatty acid concentrations and cardiac fatty acid and glucose metabolism, the use of thiazolidinediones in any cardiovascular disease state requires additional research and trials of safety.

PPAR δ is not as well characterized as PPAR α and PPAR γ ; however, experimental studies do implicate it in the regulation of fatty acid metabolism in skeletal muscle and adipose tissue. The activation of PPAR δ in both skeletal muscle [90] and adipose tissue [91] increases fatty acid β -oxidation, and thus has the potential to reduce the circulating concentration of free fatty acids to which the heart is exposed, and thereby decreases cardiac fatty acid oxidation, which has protective effects in cardiac ischemia.

Nicotinic acid

Nicotinic acid is a broad-spectrum lipid-modifying agent that possesses antiatherogenic properties, including the ability to decrease the circulating concentrations of very-low density lipoproteins and low-density lipoproteins while increasing those of high-density lipoproteins. With regards to ischemic heart disease, nicotinic acid (both as monotherapy and in combination with other lipid-decreasing drugs) has been shown to decrease the progression of atherosclerotic lesions, and to increase plaque regression [92], effects shown to decrease cardiovascular mortality (for review see [93]). In addition to its antiatherogenic properties, nicotinic acid also has the ability to modify energy substrate metabolism.

A high-affinity G-protein-coupled receptor for nicotinic acid is highly expressed in adipose tissue [94] and is most probably responsible for the unique distribution of nicotinic acid to this tissue after administration. Nicotinic acid inhibits adipose tissue lipolysis and thus decreases circulating fatty acid concentrations (*Figure 2*). These effects alter both whole-body and cardiac energy metabolism by reducing the availability of fatty acids to peripheral tissues (eg, skeletal and cardiac muscle). Human studies have demonstrated that nicotinic acid increases the cardiac respiratory quotient while not affecting the oxygen extraction ratio – effects indicative of a shift in myocardial energy substrate preference from fatty acid to carbohydrate [95,96], which may contribute to potential anti-ischemic properties in the myocardium.

Summary

The modulation of myocardial energy substrate metabolism, particularly by shifting energy substrate preference from the use of fatty acids towards the use of glucose as an oxidative fuel, is a novel therapeutic intervention, not only for angina, but also for various other manifestations of ischemic heart disease. The shift in energy substrate preference can be achieved through the use of pharmacological agents that act at several levels of the pathways of fatty acid and glucose metabolism, altering the balance and contribution of these pathways to overall cardiac energetics and thereby increasing the efficiency of both the production and utilization of ATP. Such effects can be attained by regulating the rates of flux through the pathways of fatty acid oxidation and glucose oxidation, both by manipulating the activities of key enzymes and by altering the availability of circulating substrates. The efficacy of, for example, trimetazidine in the widespread treatment of angina is of particular relevance to the feasibility of modulating and optimizing energy substrate metabolism to limit

cardiac dysfunction in the setting of ischemic heart disease. ■

REFERENCES

- Lopaschuk GD, Belke DD, Gamble J, Itoi T, Schönekeess BO. Regulation of fatty acid oxidation in the mammalian heart in health and disease. *Biochim Biophys Acta*. 1994;1213:263–276.
- Stanley WC, Chandler MP. Energy metabolism in the normal and failing heart: potential for therapeutic interventions. *Heart Fail Rev*. 2002;7:115–130.
- Young LH, Coven DL, Russell RR 3rd. Cellular and molecular regulation of cardiac glucose transport. *J Nucl Cardiol*. 2000;7:267–276.
- Holness MJ, Sugden MC. Regulation of pyruvate dehydrogenase complex activity by reversible phosphorylation. *Biochem Soc Trans*. 2003;31:1143–1151.
- Sugden MC, Holness MJ. Recent advances in mechanisms regulating glucose oxidation at the level of the pyruvate dehydrogenase complex by PDKs. *Am J Physiol Endocrinol Metab*. 2003;284:E855–E862.
- Spriet LL, Heigenhauser GJ. Regulation of pyruvate dehydrogenase (PDH) activity in human skeletal muscle during exercise. *Exerc Sport Sci Rev*. 2002;30:91–95.
- Randle PJ, Garland PB, Hales CN, Newsholme EA. The glucose fatty-acid cycle. Its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus. *Lancet*. 1963;1:785–789.
- Glatz JF, Luiken JJ, Bonen A. Involvement of membrane-associated proteins in the acute regulation of cellular fatty acid uptake. *J Mol Neurosci*. 2001;16:123–132; discussion 151–157.
- Kerner J, Hoppel C. Fatty acid import into mitochondria. *Biochim Biophys Acta*. 2000;1486:1–17.
- Mjos OD, Ichihara K, Fellenius E, Myrmed T, Neely JR. Fatty acids suppress recovery of heart function after hypothermic perfusion. *Ann Thorac Surg*. 1991;52:965–970.
- Kennedy JA, Kiosoglous AJ, Murphy GA, Pelle MA, Horowitz JD. Effect of perhexiline and oxfenicine on myocardial function and metabolism during low-flow ischemia/reperfusion in the isolated rat heart. *J Cardiovasc Pharmacol*. 2000;36:794–801.
- Lopaschuk GD, Wall SR, Olley PM, Davies NJ. Etomoxir, a carnitine palmitoyltransferase I inhibitor, protects hearts from fatty acid-induced ischemic injury independent of changes in long chain acylcarnitine. *Circ Res*. 1988;63:1036–1043.
- Lopaschuk GD, Spafford MA, Davies NJ, Wall SR. Glucose and palmitate oxidation in isolated working rat hearts reperfused after a period of transient global ischemia. *Circ Res*. 1990;66:546–553.
- Jeffrey FM, Alvarez L, Diczku V, Sherry AD, Malloy CR. Direct evidence that perhexiline modifies myocardial substrate utilization from fatty acids to lactate. *J Cardiovasc Pharmacol*. 1995;25:469–472.
- Lee L, Horowitz J, Frenneaux M. Metabolic manipulation in ischaemic heart disease, a novel approach to treatment. *Eur Heart J*. 2004;25:634–641.
- Deschamps D, DeBeco V, Fisch C, Fromenty B, Guillouzo A, Pessayre D. Inhibition by perhexiline of oxidative phosphorylation and the beta-oxidation of fatty acids: possible role in pseudoalcoholic liver lesions. *Hepatology*. 1994;19:948–961.
- Kennedy JA, Unger SA, Horowitz JD. Inhibition of carnitine palmitoyltransferase-I in rat heart and liver by perhexiline and amiodarone. *Biochem Pharmacol*. 1996;52:273–280.
- Cole PL, Beamer AD, McGowan N, et al. Efficacy and safety of perhexiline maleate in refractory angina. A double-blind placebo-controlled clinical trial of a novel antianginal agent. *Circulation*. 1990;81:1260–1270.
- Unger SA, Robinson MA, Horowitz JD. Perhexiline improves symptomatic status in elderly patients with severe aortic stenosis. *Aust N Z J Med*. 1997;27:24–28.
- Lee L, Campbell R, Scheuermann-Freestone M, et al. Metabolic modulation with perhexiline in chronic heart failure: a randomized, controlled trial of short-term use of a novel treatment. *Circulation*. 2005;112:3280–3288.
- Wallhaus TR, Taylor M, DeGrado TR, et al. Myocardial free fatty acid and glucose use after carvedilol treatment in patients with congestive heart failure. *Circulation*. 2001;103:2441–2446.
- Igarashi N, Nozawa T, Fujii N, et al. Influence of beta-adrenoceptor blockade on the myocardial accumulation of fatty acid tracer and its intracellular metabolism in the heart after ischemia-reperfusion injury. *Circ J*. 2006;70:1509–1514.
- Eichhorn EJ, Bedotto JB, Malloy CR, et al. Effect of beta-adrenergic blockade on myocardial function and energetics in congestive heart failure. Improvements in hemodynamic, contractile, and diastolic performance with bucindolol. *Circulation*. 1990;82:473–483.
- Eichhorn EJ, Heesch CM, Barnett JH, et al. Effect of metoprolol on myocardial function and energetics in patients with non-ischemic dilated cardiomyopathy: a randomized, double-blind, placebo-controlled study. *J Am Coll Cardiol*. 1994;24:1310–1320.
- Panchal AR, Stanley WC, Kerner J, Sabbah HN. Beta-receptor blockade decreases carnitine palmitoyl transferase I activity in dogs with heart failure. *J Card Fail*. 1998;4:121–126.
- Podbregar M, Voga G. Effect of selective and nonselective beta-blockers on resting energy production rate and total body substrate utilization in chronic heart failure. *J Card Fail*. 2002;8:369–378.
- Lopaschuk GD, Barr R, Thomas PD, Dyck JR. Beneficial effects of trimetazidine in ex vivo working ischemic hearts are due to a stimulation of glucose oxidation secondary to inhibition of long-chain 3-ketoacyl coenzyme a thiolase. *Circ Res*. 2003;93:e33–e37.
- Kantor PF, Lucien A, Kozak R, Lopaschuk GD. The antianginal drug trimetazidine shifts cardiac energy metabolism from fatty acid oxidation to glucose oxidation by inhibiting mitochondrial long-chain 3-ketoacyl coenzyme A thiolase. *Circ Res*. 2000;86:580–588.
- Parang P, Singh B, Arora R. Metabolic modulators for chronic cardiac ischemia. *J Cardiovasc Pharmacol Ther*. 2005;10:217–223.
- Boucher FR, Hearse DJ, Opie LH. Effects of trimetazidine on ischemic contracture in isolated perfused rat hearts. *J Cardiovasc Pharmacol*. 1994;24:45–49.
- Ruixing Y, Wenwu L, Al-Ghazali R. Trimetazidine inhibits cardiomyocyte apoptosis in a rabbit model of ischemia-reperfusion. *Transl Res*. 2007;149:152–160.
- Monti LD, Allibardi S, Piatti PM, et al. Triglycerides impair postischemic recovery in isolated hearts: roles of endothelin-1 and trimetazidine. *Am J Physiol Heart Circ Physiol*. 2001;281:H1122–H1130.
- Renaud JF. Internal pH, Na⁺, and Ca²⁺ regulation by trimetazidine during cardiac cell acidosis. *Cardiovasc Drugs Ther*. 1988;1:677–686.
- Clanachan AS. Contribution of protons to post-ischemic Na⁽⁺⁾ and Ca⁽²⁺⁾ overload and left ventricular mechanical dysfunction. *J Cardiovasc Electrophysiol*. 2006;17 (suppl 1):S141–S148.
- Ciapponi A, Pizarro R, Harrison J. Trimetazidine for stable angina. The Cochrane Database of Systematic Reviews. Available in The Cochrane Library [database on disk and CD ROM]. Updated quarterly. The Cochrane Collaboration. Oxford: Oxford Update Software, 2005; CD003614.
- Papadopoulos CL, Kanonidis IE, Kotridis PS, et al. The effect of trimetazidine on reperfusion arrhythmias in acute myocardial infarction. *Int J Cardiol*. 1996;55:137–142.
- Steg PG, Grollier G, Gallay P, et al., for the LIST Study Group. A randomized double-blind trial of intravenous trimetazidine as adjunctive therapy to primary angioplasty for acute myocardial infarction. *Int J Cardiol*. 2001;77:263–273.
- Fragasso G, Pallosi A, Puccetti P, et al. A randomized clinical trial of trimetazidine, a partial free fatty acid oxidation inhibitor, in patients with heart failure. *J Am Coll Cardiol*. 2006;48:992–998.
- Fragasso G, Perseghin G, De Cobelli F, et al. Effects of metabolic modulation by trimetazidine on left ventricular function and phosphocreatine/adenosine triphosphate ratio in patients with heart failure. *Eur Heart J*. 2006;27:942–948.
- Scirica BM. Ranolazine in patients with coronary artery disease. *Expert Opin Pharmacother*. 2007;8:2149–2157.
- Gralinski MR, Black SC, Kilgore KS, Chou AY, McCormack JG, Lucchesi BR. Cardioprotective effects of ranolazine (RS-43285) in the isolated perfused rabbit heart. *Cardiovasc Res*. 1994;28:1231–1237.

Basic article

Metabolic modulation for ischemic heart disease

42. Gralinski MR, Chi L, Park JL, et al. Protective effects of ranolazine on ventricular fibrillation induced by activation of the ATP-dependent potassium channel in the rabbit heart. *J Cardiovasc Pharmacol Ther.* 1996;1:141–148.
43. Hale SL, Kloner RA. Ranolazine, an inhibitor of the late sodium channel current, reduces postischemic myocardial dysfunction in the rabbit. *J Cardiovasc Pharmacol Ther.* 2006;11:249–255.
44. Hale SL, Leeka JA, Kloner RA. Improved left ventricular function and reduced necrosis after myocardial ischemia/reperfusion in rabbits treated with ranolazine, an inhibitor of the late sodium channel. *J Pharmacol Exp Ther.* 2006;318:418–423.
45. Chandler MP, Stanley WC, Morita H, et al. Short-term treatment with ranolazine improves mechanical efficiency in dogs with chronic heart failure. *Circ Res.* 2002;91:278–280.
46. Sabbah HN, Chandler MP, Mishima T, et al. Ranolazine, a partial fatty acid oxidation (pFOX) inhibitor, improves left ventricular function in dogs with chronic heart failure. *J Card Fail.* 2002;8:416–422.
47. Clarke B, Spedding M, Patmore L, McCormack JG. Protective effects of ranolazine in guinea-pig hearts during low-flow ischaemia and their association with increases in active pyruvate dehydrogenase. *Br J Pharmacol.* 1993;109:748–750.
48. Clarke B, Wyatt KM, McCormack JG. Ranolazine increases active pyruvate dehydrogenase in perfused normoxic rat hearts: evidence for an indirect mechanism. *J Mol Cell Cardiol.* 1996;28:341–350.
49. McCormack JG, Barr RL, Wolff AA, Lopaschuk GD. Ranolazine stimulates glucose oxidation in normoxic, ischemic, and reperfused ischemic rat hearts. *Circulation.* 1996;93:135–142.
50. Chaitman BR, Skettino SL, Parker JO, et al., for the MARISA Investigators. Anti-ischemic effects and long-term survival during ranolazine monotherapy in patients with chronic severe angina. *J Am Coll Cardiol.* 2004;43:1375–1382.
51. Chaitman BR, Pepine CJ, Parker JO, et al., for the Combination Assessment of Ranolazine In Stable Angina (CARISA) Investigators. Effects of ranolazine with atenolol, amlodipine, or diltiazem on exercise tolerance and angina frequency in patients with severe chronic angina: a randomized controlled trial. *JAMA.* 2004;291:309–316.
52. Rousseau MF, Pouleur H, Cocco G, Wolff AA. Comparative efficacy of ranolazine versus atenolol for chronic angina pectoris. *Am J Cardiol.* 2005;95:311–316.
53. Stone PH, Gratsiansky NA, Blokhin A, et al., for the ERICA Investigators. Antianginal efficacy of ranolazine when added to treatment with amlodipine: the ERICA (Efficacy of Ranolazine in Chronic Angina) trial. *J Am Coll Cardiol.* 2006;48:566–575.
54. Timmis AD, Chaitman BR, Crager M. Effects of ranolazine on exercise tolerance and HbA1c in patients with chronic angina and diabetes. *Eur Heart J.* 2006;27:42–48.
55. Scirica BM, Morrow DA, Hod H, et al. Effect of ranolazine, an antianginal agent with novel electrophysiological properties, on the incidence of arrhythmias in patients with non ST-segment elevation acute coronary syndrome: results from the Metabolic Efficiency With Ranolazine for Less Ischemia in Non ST-Elevation Acute Coronary Syndrome Thrombolysis in Myocardial Infarction 36 (MERLIN-TIMI 36) randomized controlled trial. *Circulation.* 2007;116:1647–1652.
56. Morrow DA, Scirica BM, Karwatowska-Prokopczuk E, et al., for the MERLIN-TIMI 36 Trial Investigators. Effects of ranolazine on recurrent cardiovascular events in patients with non-ST-elevation acute coronary syndromes: the MERLIN-TIMI 36 randomized trial. *JAMA.* 2007;297:1775–1783.
57. Belardinelli L, Shryock J, Fraser H. Inhibition of the late sodium current as a potential cardioprotective principle: effects of the late sodium current inhibitor ranolazine. *Heart.* 2006;92 (suppl 4):iv6–iv14.
58. McVeigh JJ, Lopaschuk GD. Dichloroacetate stimulation of glucose oxidation improves recovery of ischemic rat hearts. *Am J Physiol Heart Circ Physiol.* 1990;259:H1079–H1085.
59. Itoi T, Huang L, Lopaschuk GD. Glucose use in neonatal rabbit hearts reperfused after global ischemia. *Am J Physiol Heart Circ Physiol.* 1993;265:H427–H433.
60. Stanley WC, Hernandez LA, Spires D, Bringas J, Wallace S, McCormack JG. Pyruvate dehydrogenase activity and malonyl CoA levels in normal and ischemic swine myocardium: effects of dichloroacetate. *J Mol Cell Cardiol.* 1996;28:905–914.
61. Liu B, Clanachan AS, Schulz R, Lopaschuk GD. Cardiac efficiency is improved after ischemia by altering both the source and fate of protons. *Circ Res.* 1996;79:940–948.
62. Liu Q, Docherty JC, Rendell JC, Clanachan AS, Lopaschuk GD. High levels of fatty acids delay the recovery of intracellular pH and cardiac efficiency in post-ischemic hearts by inhibiting glucose oxidation. *J Am Coll Cardiol.* 2002;39:718–725.
63. Wargovich TJ, MacDonald RG, Hill JA, Feldman RL, Stac-pool PW, Pepine CJ. Myocardial metabolic and hemodynamic effects of dichloroacetate in coronary artery disease. *Am J Cardiol.* 1988;61:65–70.
64. Sodi-Pallares D, Testelli MR, Fishleder BL, et al. Effects of an intravenous infusion of a potassium–glucose–insulin solution on the electrocardiographic signs of myocardial infarction. A preliminary clinical report. *Am J Cardiol.* 1962;9:166–181.
65. Opie LH. The glucose hypothesis: relation to acute myocardial ischaemia. *J Mol Cell Cardiol.* 1970;1:107–115.
66. Jonassen AK, Aasum E, Riemersma RA, Mjøs OD, Larsen TS. Glucose–insulin–potassium reduces infarct size when administered during reperfusion. *Cardiovasc Drugs Ther.* 2000;14:615–623.
67. Zhang HX, Zang YM, Huo JH, et al. Physiologically tolerable insulin reduces myocardial injury and improves cardiac functional recovery in myocardial ischemic/reperfused dogs. *J Cardiovasc Pharmacol.* 2006;48:306–313.
68. Kloner RA, Przyklenk K, Shook T, Cannon CP. Protection conferred by preinfarct angina is manifest in the aged heart: evidence from the TIMI 4 Trial. *J Thromb Thrombolysis.* 1998;6:89–92.
69. Folmes CD, Clanachan AS, Lopaschuk GD. Fatty acids attenuate insulin regulation of 5'-AMP-activated protein kinase and insulin cardioprotection after ischemia. *Circ Res.* 2006;99:61–68.
70. Fath-Ordoubadi F, Beatt KJ. Glucose–insulin–potassium therapy for treatment of acute myocardial infarction: an overview of randomized placebo-controlled trials. *Circulation.* 1997;96:1152–1156.
71. Malmberg K, Rydén L, Hamsten A, Herlitz J, Waldenström A, Wedel H. Effects of insulin treatment on cause-specific one-year mortality and morbidity in diabetic patients with acute myocardial infarction. DIGAMI Study Group. *Diabetes Insulin-Glucose in Acute Myocardial Infarction. Eur Heart J.* 1996;17:1337–1344.
72. Diaz R, Paolasso EA, Piegas LS, et al. Metabolic modulation of acute myocardial infarction. The ECLA (Estudios Cardiologicos Latinoamericana) Collaborative Group. *Circulation.* 1998;98:2227–2234.
73. Ceremuzynski L, Budaj A, Czepiel A, et al. Low-dose glucose–insulin–potassium is ineffective in acute myocardial infarction: results of a randomized multicenter Pol-GIK trial. *Cardiovasc Drugs Ther.* 1999;13:191–200.
74. van der Horst IC, Zijlstra F, van 't Hof AW, et al., for the Zwolle Infarct Study Group. Glucose–insulin–potassium infusion in patients treated with primary angioplasty for acute myocardial infarction: the glucose–insulin–potassium study: a randomized trial. *J Am Coll Cardiol.* 2003;42:784–791.
75. Timmer J, GIPS 2 Investigators. Glucose–Insulin–Potassium Study in patients with ST-segment elevation myocardial infarction without signs of heart failure. In: Late Breaking Clinical Trials III. American College of Cardiology Scientific Sessions; 2005; Orlando.
76. Smith SA. Peroxisome proliferator-activated receptors and the regulation of mammalian lipid metabolism. *Biochem Soc Trans.* 2002;30:1086–1090.
77. Gilde AJ, Van Bilsen M. Peroxisome proliferator-activated receptors (PPARs): regulators of gene expression in heart and skeletal muscle. *Acta Physiol Scand.* 2003;178:425–434.
78. Luquet S, Lopez-Soriano J, Holst D, et al. Roles of peroxisome proliferator-activated receptor delta (PPARdelta) in the control of fatty acid catabolism. A new target for the treatment of metabolic syndrome. *Biochimie.* 2004;86:833–837.
79. Paulussen RJ, Jansen GP, Veerkamp JH. Fatty acid-binding capacity of cytosolic proteins of various rat tissues: effect of postnatal development, starvation, sex, clofibrate feeding and light cycle. *Biochim Biophys Acta.* 1986;877:342–349.
80. Schoonjans K, Staels B, Grimaldi P, Auwerx J. Acyl-CoA synthetase mRNA expression is controlled by fibric-acid derivatives, feeding and liver proliferation. *Eur J Biochem.* 1993;216:615–622.

Basic article

Jagdip S. Jaswal, Virgilio J. J. Cadete and Gary D. Lopaschuk

81. Cook WS, Yeldandi AV, Rao MS, Hashimoto T, Reddy JK. Less extrahepatic induction of fatty acid beta-oxidation enzymes by PPAR alpha. *Biochem Biophys Res Commun.* 2000;278:250–257.
82. Prasad MR, Clement R, Otani H, et al. Improved myocardial performance induced by clofibrate during reperfusion after acute myocardial infarction. *Can J Physiol Pharmacol.* 1988;66:1518–1523.
83. Wayman NS, Hattori Y, McDonald MC, et al. Ligands of the peroxisome proliferator-activated receptors (PPAR-gamma and PPAR-alpha) reduce myocardial infarct size. *FASEB J.* 2002;16:1027–1040.
84. Yue TL, Bao W, Gu JL, et al. Rosiglitazone treatment in Zucker diabetic fatty rats is associated with ameliorated cardiac insulin resistance and protection from ischemia/reperfusion-induced myocardial injury. *Diabetes.* 2005;54:554–562.
85. Zhu P, Lu L, Xu Y, Schwartz GG. Troglitazone improves recovery of left ventricular function after regional ischemia in pigs. *Circulation.* 2000;101:1165–1171.
86. Sidell RJ, Cole MA, Draper NJ, Desrois M, Buckingham RE, Clarke K. Thiazolidinedione treatment normalizes insulin resistance and ischemic injury in the Zucker fatty rat heart. *Diabetes.* 2002;51:1110–1117.
87. Yue TL, Chen J, Bao W, et al. In vivo myocardial protection from ischemia/reperfusion injury by the peroxisome proliferator-activated receptor-gamma agonist rosiglitazone. *Circulation.* 2001;104:2588–2594.
88. Lindenfeld J, Masoudi FA. Fluid retention with thiazolidinediones: does the mechanism influence the outcome? *J Am Coll Cardiol.* 2007;49:1705–1707.
89. Nissen SE, Wolski K. Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. *N Engl J Med.* 2007;356:2457–2471.
90. Tanaka T, Yamamoto J, Iwasaki S, et al. Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-oxidation in skeletal muscle and attenuates metabolic syndrome. *Proc Natl Acad Sci U S A.* 2003;100:15924–15929.
91. Wang YX, Lee CH, Tiep S, et al. Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell.* 2003;113:159–170.
92. McKenney JM, Jones PH, Bays HE, et al. Comparative effects on lipid levels of combination therapy with a statin and extended-release niacin or ezetimibe versus a statin alone (the COMPELL study). *Atherosclerosis.* 2007;192:432–437.
93. Carlson LA. Nicotinic acid: the broad-spectrum lipid drug. A 50th anniversary review. *J Intern Med.* 2005;258:94–114.
94. Tunaru S, Kero J, Schaub A, et al. PUMA-G and HM74 are receptors for nicotinic acid and mediate its anti-lipolytic effect. *Nat Med.* 2003;9:352–355.
95. Carlson LA, Lassers BW, Wahlqvist ML, Kaijser L. The relationship in man between plasma free fatty acids and myocardial metabolism of carbohydrate substrates. *Cardiology.* 1972;57:51–54.
96. Lassers BW, Wahlqvist ML, Kaijser L, Carlson LA. Effect of nicotinic acid on myocardial metabolism in man at rest and during exercise. *J Appl Physiol.* 1972;33:72–80.